Cat. # RR420A

For Research Use

TaKaRa

TB Green® *Premix Ex Taq*™ (Tli RNaseH Plus)

Product Manual

v202607w

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I. Description

TB Green *Premix Ex Taq* (Tli RNaseH Plus) is designed for intercalator-based real-time PCR using TB Green. It is supplied at a 2X concentration and includes TB Green at a concentration appropriate for real time monitoring, making it easy to prepare reaction mixtures. The 2X premixed reagent also contains Tli RNaseH, a heat-resistant RNase H that minimizes PCR inhibition by degrading residual mRNA when using cDNA as template.

The combination of *TaKaRa Ex Taq*® HS, a hot-start PCR enzyme that includes an anti-*Taq* antibody, and a buffer optimized for real-time PCR, allows high amplification efficiency and detection sensitivity. This product is suitable for high-speed PCR and enables accurate assay and detection of targets, making it possible to obtain highly reproducible and reliable real-time PCR results.

Benefits

(1) Allows rapid and accurate detection and assay of targets by real-time PCR.

(2) 2X concentration premixed with TB Green; simply add primers, template, and sterile purified water to perform intercalator-based real-time PCR.

(3) *TaKaRa Ex Taq* HS, a hot-start PCR enzyme, is used for PCR. The buffer system has been optimized for real-time PCR, allowing good amplification efficiency and high-sensitivity detection.

(4) The 2X reagent is premixed with Tli RNaseH, a heat-resistant RNase H that minimizes PCR inhibition by residual mRNA when using cDNA as template.

II. Principle

This product uses *TaKaRa Ex Taq* HS for PCR amplification. PCR amplification products may be monitored in real time using TB Green as an intercalator.

1. PCR

PCR is a technique used to amplify specific target sequences from minute amounts of DNA. By repeating three cycles of heat denaturation, primer annealing, and primer extension, the target fragment is amplified up to a million times by DNA polymerase within a short time.

This product uses *TaKaRa Ex Taq* HS, a hot-start PCR enzyme that prevents non-specific amplification resulting from mispriming or primer dimer formation during reaction mixture preparation or other pre-cycling steps thereby allowing high-sensitivity detection.

2. Fluorescent detection - Intercalator method

This method uses a DNA intercalator (e.g., TB Green) that emits fluorescence when bound to double-strand DNA. Monitoring fluorescence allows for quantification of amplification products.

Measuring the fluorescence intensity also provides the melting temperature of amplified DNA.

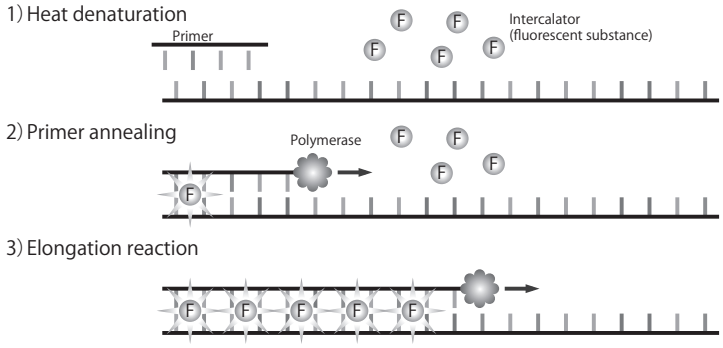


Figure 1. Fluorescent intercalator detection method

III. Components (200 reactions, 50 μl volume)

TB Green *Premix Ex Taq* (Tli RNaseH Plus) (2X conc.)*1	1 ml x 5
ROX Reference Dye (50X conc.)*2	200 μl
ROX Reference Dye II (50X conc.)*2	200 μl

*1 Contains *TaKaRa Ex Taq* HS, dNTP Mixture, Mg^{2+}, Tli RNaseH, and TB Green.

*2 ROX Reference Dye is used for analyses with devices that correct for between-well fluorescent signal, such as the real-time PCR devices by Applied Biosystems .

◆ Use ROX Reference Dye
- Applied Biosystems 7300 Real-Time PCR System
- Applied Biosystems StepOnePlus Real-Time PCR System
(Both models are made by Thermo Fisher Scientific.)

◆ Use ROX Reference Dye II
- Applied Biosystems QuantStudio 3/5 Real-Time PCR System
- Applied Biosystems 7500/7500 Fast Real-Time PCR System
(These models are made by Thermo Fisher Scientific.)

◆ Do not use this component
- Thermal Cycler Dice™ Real Time System series
- Smart Cycler II System (Cepheid)
- LightCycler/LightCycler 480 System (Roche Diagnostics)
- CFX96 Real-Time PCR Detection System (Bio-Rad)

IV. Materials Required but not Provided

1. Reagents

- PCR primers
  For real-time PCR primer design, please refer to Section IX.-1.
- Sterile purified water

2. Materials

- Real-time PCR reaction tubes or plates
- Micropipettes and tips (autoclave treated)
- Gene amplification system for real-time PCR (authorized instruments)

  Compatible instruments include:
  - Applied Biosystems QuantStudio 3/5 Real-Time PCR System
    Applied Biosystems 7300/7500/7500 Fast Real-Time PCR System
    Applied Biosystems StepOnePlus Real-Time PCR System
    (These models are made by Thermo Fisher Scientific.)
  - LightCycler/LightCycler 480 System (Roche Diagnostics)
  - Smart Cycler II System (Cepheid)
  - CFX96 Real-Time PCR Detection System (Bio-Rad)
  - Thermal Cycler Dice Real Time System IV (Cat. #TP1000/TP1010/TP1030)*
    Thermal Cycler Dice Real Time System III (Cat. #TP950/TP970/TP980/TP990)*

  * Not available in all geographic locations. Check for availability in your area.

V. Storage

Store at 4℃ (stable for up to 6 months.)

Always protect from light and avoid contamination.

For long-term storage, store at -20℃. Store thawed or opened product at 4℃ and use within 6 months.

VI. Precautions

Read these precautions before use and follow them when using this product.

1. Before use, make sure the reagent is evenly mixed by gently inverting several times without creating bubbles. Uneven reagent mixing will result in inadequate reactivity.
   - Do not mix by vortexing.
   - When stored at -20℃, TB Green *Premix Ex Taq* (2X conc.) may develop a white to pale yellow precipitant. Gently hand-warm and let stand protected from light at room temperature. Invert gently several times to dissolve the precipitate completely.
   - The presence of precipitant is indicative of uneven reagent distribution; make sure that the reagent is evenly mixed before use.
2. Place reagents on ice when preparing the reaction mixture.

3. This product contains TB Green. Avoid exposure to strong light when preparing the reaction mixture.
4. Use fresh disposable tips to avoid contamination between samples when preparing or dispensing reaction mixtures.

VII. Protocol

Note: Please follow the procedures outlined in the manual of each respective instrument.

【Applied Biosystems QuantStudio 3/5 Real-Time PCR System, 7300/7500/7500 Fast Real-Time PCR System, and StepOnePlus Real-Time PCR System】

A. Prepare the PCR mixture shown below.

<Per reaction>

Reagent	Volume	Volume	Final conc.
TB Green *Premix Ex Taq* (Tli RNaseH Plus) (2X)	10 μl	25 μl	1X
PCR Forward Primer (10 μM)	0.4 μl	1 μl	0.2 μM*1
PCR Reverse Primer (10 μM)	0.4 μl	1 μl	0.2 μM*1
ROX Reference Dye (50X) or Dye II (50X)*2	0.4 μl	1 μl	1X
Template*3	2 μl	4 μl	
Sterile purified water	6.8 μl	18 μl	
Total	20 μl*4	50 μl*4	

* 1 A final primer concentration of 0.2 μM likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and 1.0 μM.

*2 The concentration of ROX Reference Dye II (50X) is lower than that of ROX Reference Dye (50X).

- Use ROX Reference Dye II (50X) when performing analyses with Applied Biosystems QuantStudio 3/5 Real-Time PCR System or 7500/7500 Fast Real-Time PCR System.
- Use ROX Reference Dye (50X) when using Applied Biosystems StepOnePlus or Applied Biosystems 7300 Real-Time PCR System.

*3 The quantity varies depending on the number of target copies present in the template solution. Make serial dilutions to determine the appropriate template amount and use no more than 100 ng of DNA template per 20 μl. Furthermore, if cDNA (RT reaction mixture) is used as template, the volume of the RT reaction mixture should be no more than 10% of PCR mixture.

*4 Prepare in accordance with the recommended volume for each instrument.

B. Start the reaction.

The recommended two-step PCR protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform three-step PCR when using primers with low Tm values or when a two-step PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)

1) Applied Biosystems QuantStudio 3/5 Real-Time PCR System



Two-step PCR standard protocol

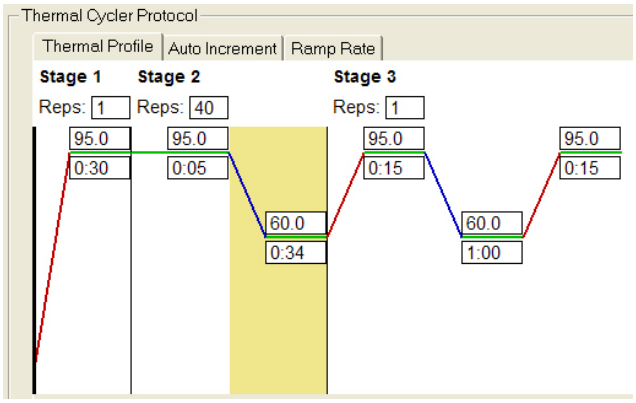
Hold (Initial Denaturation)
95℃ 30 sec

PCR
Number of Cycles: 40
95℃ 5 sec
60℃ 30 sec

Melt Curve

Figure 2. Two-step PCR standard protocol

2) Applied Biosystems 7300/7500 and StepOnePlus Real-Time PCR System



Stage 1: Initial denaturation
Reps: 1
95℃ 30 sec

Stage 2: PCR
Reps: 40
95℃ 5 sec
60℃ 30 - 34 sec *

Dissociation stage

* StepOnePlus, set to 30 sec;
7300, set to 31 sec;
7500, set to 34 sec

Figure 3. Two-step PCR standard protocol

3) Applied Biosystems 7500 Fast Real-Time PCR System

Two-step PCR standard protocol

Hold Stage
Number of Cycle: 1
95℃ 30 sec
Cycling Stage
Number of Cycles: 40
95℃ 3 sec
60℃ 30 sec
Melt Curve Stage

Note:

- *TaKaRa Ex Taq* HS is a hot-start PCR enzyme that includes an anti-*Taq* antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95℃ for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95℃ for 30 sec is sufficient.

C. After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed.

Refer to the instrument's instruction manual for specific analysis methods.

【 LightCycler/LightCycler 480 System 】

A. Prepare the PCR mixture shown below.

<Per reaction>

Reagent	Volume	Final conc.
TB Green *Premix Ex Taq* (Tli RNaseH Plus) (2X)	10 μl	1X
PCR Forward Primer (10 μM)	0.4 μl	0.2 μM*1
PCR Reverse Primer (10 μM)	0.4 μl	0.2 μM*1
Template (< 100 ng)*2	2 μl	
Sterile purified water	7.2 μl	
Total	20 μl	

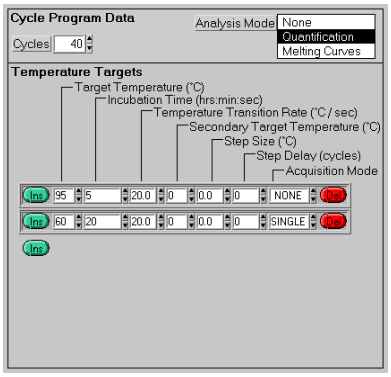
*1 A final primer concentration of 0.2 μM likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and 1.0 μM.

*2 The optimal amount depends on the number of target copies in the template solution. Make serial dilutions to determine the appropriate amount and to use no more than 100 ng of DNA template. Furthermore, if cDNA (RT reaction mixture) is added as template, the template volume should be no more than 10% of the PCR mixture.

B. Start the reaction.

The recommended two-step PCR protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform three-step PCR when using primers with low Tm values or when a two-step PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)

<LightCycler>



Stage 1: Initial denaturation
95℃ 30 sec 20℃/sec
1 cycle

Stage 2: PCR (See figure on the left)
95℃ 5 sec 20℃/sec
60℃ 20 sec 20℃/sec
40 cycles

Stage 3: Melt Curve Analysis
95℃ 0 sec 20℃/sec
65℃ 15 sec 20℃/sec
95℃ 0 sec 0.1℃/sec

Figure 4. Two-step PCR standard protocol

<LightCycler 480 System>

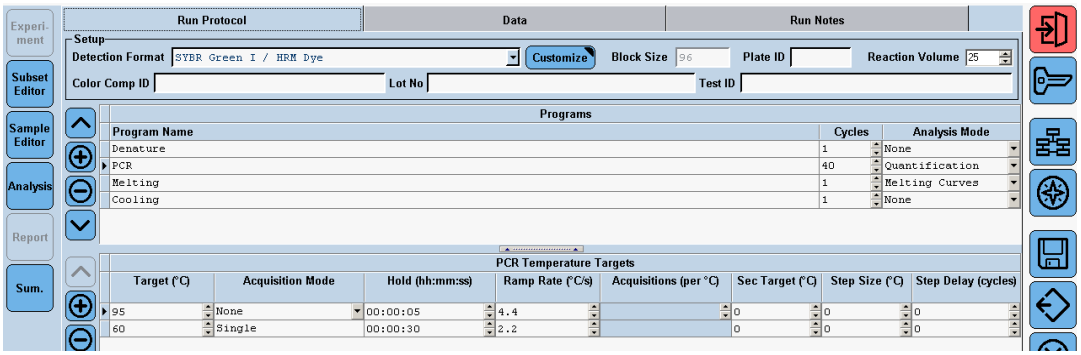


Figure 5. Two-step PCR standard protocol

Denature
95℃ 30 sec (Ramp rate: 4.4℃/sec)
1 cycle

PCR
Analysis Mode: Quantification
95℃ 5 sec (Ramp rate: 4.4℃/sec)
60℃ 30 sec (Ramp rate: 2.2℃/sec, Acquisition Mode: Single)
40 cycles

Melting
Analysis Mode: Melting Curves
95℃ 5 sec (Ramp rate: 4.4℃/sec)
60℃ 1 min (Ramp rate: 2.2℃/sec)
95℃ (Ramp rate: 0.11℃/sec, Acquisition Mode: Continuous, Acquisitions: 5 per ℃)
1 cycle

Cooling
50℃ 30 sec (Ramp rate: 2.2℃/sec)
1 cycle

Note:
- *TaKaRa Ex Taq* HS is a hot-start PCR enzyme includes an anti-*Taq* antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95℃ for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95℃ for 30 sec is sufficient.

C. After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed.
Refer to the instrument manual for specific analysis methods.

【 Smart Cycler II System 】

A. Prepare the PCR mixture shown below.

<Per reaction>

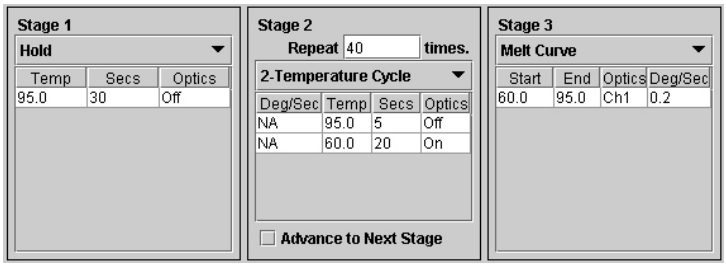
Reagent	Volume	Final conc.
TB Green *Premix Ex Taq* (Tli RNaseH Plus) (2X)	12.5 μl	1X
PCR Forward Primer (10 μM)	0.5 μl	0.2 μM*1
PCR Reverse Primer (10 μM)	0.5 μl	0.2 μM*1
Template (< 100 ng)*2	2.0 μl	
Sterile purified water	9.5 μl	
Total	25 μl	

*1 A final primer concentration of 0.2 μM likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and 1.0 μM.

*2 The quantity varies depending on the number of target copies present in the template solution. Make serial dilutions to determine the appropriate amount and use no more than 100 ng of DNA template. If cDNA (RT reaction mixture) is used as template, the volume of the RT reaction mixture should be no more than 10% of the PCR mixture.

B. Briefly centrifuge reaction tubes with the Smart Cycler centrifuge and then set them in the Smart Cycler instrument to initiate the reaction.

The recommended two-step PCR protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform three-step PCR when using primers with low Tm values or when a two-step PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)



Stage 1: Initial denaturation
Hold
95℃ 30 sec
Stage 2: PCR
Repeat: 40 times
95℃ 5 sec
60℃ 20 sec
Stage 3: Melt curve

Figure 6. Two-step PCR standard protocol

Note:

- *TaKaRa Ex Taq* HS is a hot-start PCR enzyme that includes an anti-*Taq* antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95℃ for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95℃ for 30 sec is sufficient.

C. After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed.

For the analysis methods when using the Smart Cycler System, please refer to the instruction manual for Smart Cycler System.

【 CFX96 Real-Time PCR Detection System 】

A. Prepare the PCR mixture shown below.

<Per reaction>

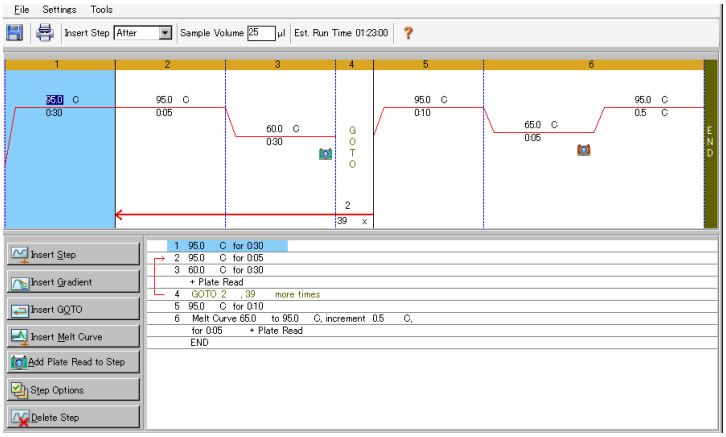
Reagent	Volume	Final conc.
TB Green *Premix Ex Taq* (Tli RNaseH Plus) (2X)	12.5 μl	1X
PCR Forward Primer (10 μM)	0.5 μl	0.2 μM*1
PCR Reverse Primer (10 μM)	0.5 μl	0.2 μM*1
Template (< 100 ng)*2	2 μl	
Sterile purified water	9.5 μl	
Total	25 μl	

*1 A final primer concentration of 0.2 μM likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and 1.0 μM.

*2 The optimal amount depends on the number of target copies in the template solution. Make serial dilutions to determine the appropriate amount and use no more than 100 ng of DNA template. If cDNA (RT reaction mixture) is added as template, the template volume should be no more than 10% of the PCR mixture.

B. Start the reaction.

The recommended two-step PCR protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform three-step PCR when using primers with low Tm values or when a two-step PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)



Sample volume: 25 μl

Step 1:
95℃ 30 sec
Step 2: PCR
GOTO: 39 (40 cycles)
95℃ 5 sec
60℃ 30 sec
Step 3: Melt Curve

Figure 7. Two-step PCR standard protocol

Note:

- *TaKaRa Ex Taq* HS is a hot-start PCR enzyme that includes an anti-*Taq* antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95℃ for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95℃ for 30 sec is sufficient.

C. After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed.

For analysis methods, refer to the manual for CFX96 Real-Time PCR Detection System.

【 Thermal Cycler Dice Real Time System Series 】

A. Prepare the PCR mixture shown below.

<Per reaction>

Reagent	Volume	Final conc.
TB Green *Premix Ex Taq* (Tli RNaseH Plus) (2X)	12.5 μl	1X
PCR Forward Primer (10 μM)	0.5 μl	0.2 μM*1
PCR Reverse Primer (10 μM)	0.5 μl	0.2 μM*1
Template (< 100 ng)*2	2.0 μl	
Sterile purified water	9.5 μl	
Total	25 μl*3	

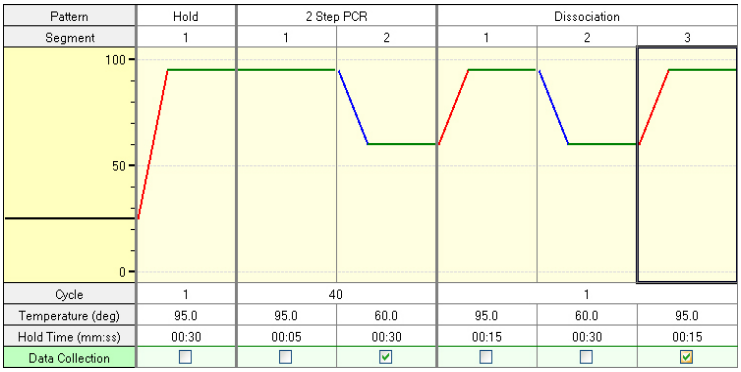
*1 A final primer concentration of 0.2 μM likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and 1.0 μM.

*2 The optimal amount varies depending on the number of target copies present in the template solution. Make serial dilutions to determine the appropriate amount and use no more than 100 ng of DNA template. If cDNA (RT reaction mixture) is used as template, the volume of the RT reaction mixture should be no more than 10% of PCR mixture.

*3 The recommended volume is 25 μl for reaction mixtures.

B. Start the reaction.

The recommended two-step PCR protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform three-step PCR when using primers with low Tm values or when a two-step PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)



Hold (initial denaturation)
Cycle: 1
95℃ 30 sec

Two-step PCR
Cycle: 40
95℃ 5 sec
60℃ 30 sec

Dissociation

Figure 8. Two-step PCR standard protocol

Note:

- *TaKaRa Ex Taq* HS is a hot-start PCR enzyme that includes an anti-*Taq* antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95℃ for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95℃ for 30 sec is sufficient.

C. After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed.
When using Thermal Cycler Dice Real Time System, please refer to the instruction manual for analysis methods.

VIII. Optimization

If unsatisfactory results are obtained using the recommended conditions (two-step PCR standard protocol), follow the procedures below to optimize the primer concentration and PCR conditions. In addition, depending on the reaction system, switching to another TB Green Premix (Cat. #RR820A/B, RR091A/B) may greatly improve the results.

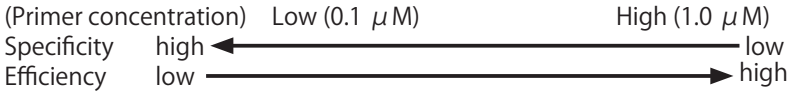
Select PCR conditions based on comprehensive analysis of both reaction specificity and amplification efficiency. Using conditions that balance these two aspects will allow accurate measurement over a wide range of concentrations.

○ System with a high reaction specificity
- With no template control, non-specific amplification (e.g., primer-dimers) does not occur.
- Non-specific amplification products, those other than the target product, are not generated.

○ System with a high amplification efficiency
- Amplification product is detected early (small Ct value).
- PCR amplification efficiency is high (near the theoretical value of 100%).

1. Evaluation of primer concentration

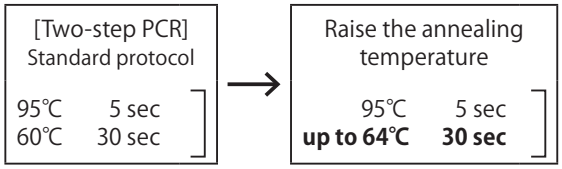
The relationship between primer concentration and reaction specificity and amplification efficiency is illustrated below. Reducing primer concentration raises reaction specificity. In contrast, increasing primer concentration raises amplification efficiency.



2. Evaluation of PCR conditions

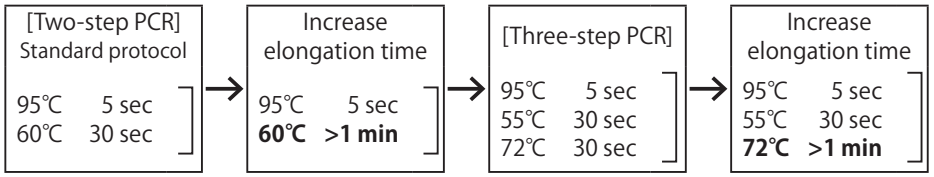
○ To improve reaction specificity

Raising the annealing temperature may improve reaction specificity. Perform optimization while checking for effects on amplification efficiency.



○ To improve amplification efficiency

Increasing elongation time or switching to a three-step PCR protocol may improve amplification efficiency. Perform optimization using the steps below.



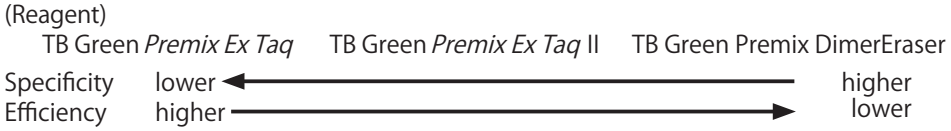
○ Initial denaturation

Generally, 95℃ for 30 sec is sufficient for initial denaturation, even for difficult-to-denature templates such as circular plasmids and genomic DNA. This step may be extended to 1 - 2 min at 95℃ depending on the template. However, prolonging this step may inactivate the enzyme. Therefore, it is recommended to avoid initial denaturation steps longer than 2 min.

3. Relationship between reagent and reactivity

Takara Bio supplies several different reagents for intercalator-based real-time PCR analysis using TB Green. The relationship between reaction specificity and amplification efficiency for these reagents is described below.

TB Green *Premix Ex Taq* (Tli RNaseH Plus) (Cat. #RR420A/B) provides high amplification efficiency. TB Green *Premix Ex Taq* II (Tli RNaseH Plus) (Cat. #RR820A/B) and TB Green Premix DimerEraser™ (Perfect Real Time) (Cat. #RR091A/B) have greater specificity.



IX. Appendix

1. Primer design

Designing primers with good reactivity is critical to efficient real-time PCR. Please follow the guidelines below to design primers that yield high amplification efficiency without non-specific amplification.

RT-PCR primers designed and synthesized using these guidelines are compatible with the standard two-step PCR protocol (Section VII.).

■ Amplification product

Amplification size	The optimal size is 80 - 150 bp (amplification up to 300 bp is possible)

■ Primer

Length	17 - 25mer
GC content	40 - 60% (preferably 45 - 55%)
Tm	Make sure that the Tm values for the forward primer and the reverse primer do not differ greatly. Use primer design software to determine Tm values. OLIGO*1 : 63 - 68℃ Primer3 : 60 - 65℃
Sequence	Make sure that overall there are no base sequence biases. Avoid having any GC-rich or AT-rich regions in the sequence (particularly at the 3′ end). Avoid having consecutive T/C pairings (polypyrimidine). Avoid having consecutive A/G pairings (polypurine).
3′ end sequence	Avoid having any GC-rich or AT-rich sequence at the 3′ end. It is preferable to have a G or C as the 3′ end-base. Avoid primers with T as the 3′ end-base.
Complementation	Avoid having any complementary sequences of 3 bases or more within a primer and between primers. Avoid having any complementary sequences of 2 bases or more at the primer's 3' ends.
Specificity	Verify primer specificity by BLAST search.*2

*1 OLIGO Primer Analysis Software (Molecular Biology Insights)

*2 https://blast.ncbi.nlm.nih.gov/Blast.cgi

2. When performing real-time RT-PCR

To synthesize cDNA templates for real-time RT-PCR, we recommend using PrimeScript™ reverse transcriptase products.

- PrimeScript RT reagent Kit (Perfect Real Time) (Cat. #RR037A/B)
- PrimeScript RT Master Mix (Perfect Real Time) (Cat. #RR036A/B)
- PrimeScript RT reagent Kit with gDNA Eraser (Perfect Real Time) (Cat. #RR047A/B)

When used in combination with this kit, these products can provide highly reliable results.

A. Prepare a PCR mixture by the following procedure.
(When using Thermal Cycler Dice Real Time System)

Prepare the following components in volumes slightly more than that needed for the required number of tubes and dispense 22.5 - 24 μl into tubes.

<Per reaction>

Reagent	Volume	Final conc.
TB Green *Premix Ex Taq* (2X)	12.5 μl	1X
PCR Forward Primer (10 μM)	0.5 μl	0.2 μM
PCR Reverse Primer (10 μM)	0.5 μl	0.2 μM
Sterile purified water	x μl	
Total	22.5 - 24 μl	

B. Add 1 - 2.5 μl of the reverse transcription reaction mixture to each of the microtubes containing aliquots of the reaction mixture.

Note: Add no more than 2.5 μl of the reverse transcription reaction solution to the PCR mixture.

[Experimental example]

Human ATP5F1 mRNA was detected by real-time RT-PCR. cDNA equivalent to 1 pg - 100 ng of total RNA was used as the template, with sterile purified water as the negative control.

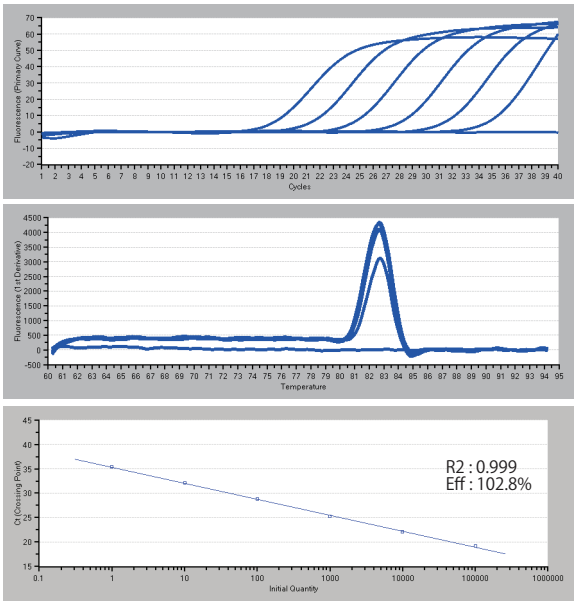


Figure 9. Detection of human ATP5F1 mRNA using real-time RT-PCR

X. Related Products

TB Green® *Premix Ex Taq*™ (Tli RNaseH Plus), Bulk (Cat. #RR420L/W)*
TB Green® *Premix Ex Taq*™ (Tli RNaseH Plus), ROX plus (Cat. #RR42LR/WR)*
TB Green® Fast qPCR Mix (Cat. #RR430A/B)
TB Green® *Premix Ex Taq*™ II (Tli RNaseH Plus) (Cat. #RR820S/A/B)*
TB Green® *Premix Ex Taq*™ II (Tli RNaseH Plus), Bulk (Cat. #RR820L/W)*
TB Green® *Premix Ex Taq*™ II (Tli RNaseH Plus), ROX plus (Cat. #RR82LR/WR)*
TB Green® Premix DimerEraser™ (Perfect Real Time) (Cat. #RR091A/B)*
TB Green® *Premix Ex Taq*™ GC (Perfect Real Time) (Cat. #RR071A/B)*
PrimeScript™ RT reagent Kit (Perfect Real Time) (Cat. #RR037A/B)
PrimeScript™ RT Master Mix (Perfect Real Time) (Cat. #RR036A/B)
PrimeScript™ RT reagent Kit with gDNA Eraser (Perfect Real Time) (Cat. #RR047A/B)
Thermal Cycler Dice™ Real Time System IV (Cat. #TP1000/TP1010/TP1030)*
Thermal Cycler Dice™ Real Time System III (Cat. #TP950/TP970/TP980/TP990)*

* Not available in all geographic locations. Check for availability in your area.

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